



Emerging Rodent Pathogens and Their Role in the Rodent Sentinel Program

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May 10, 2012



Outline

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Introduction

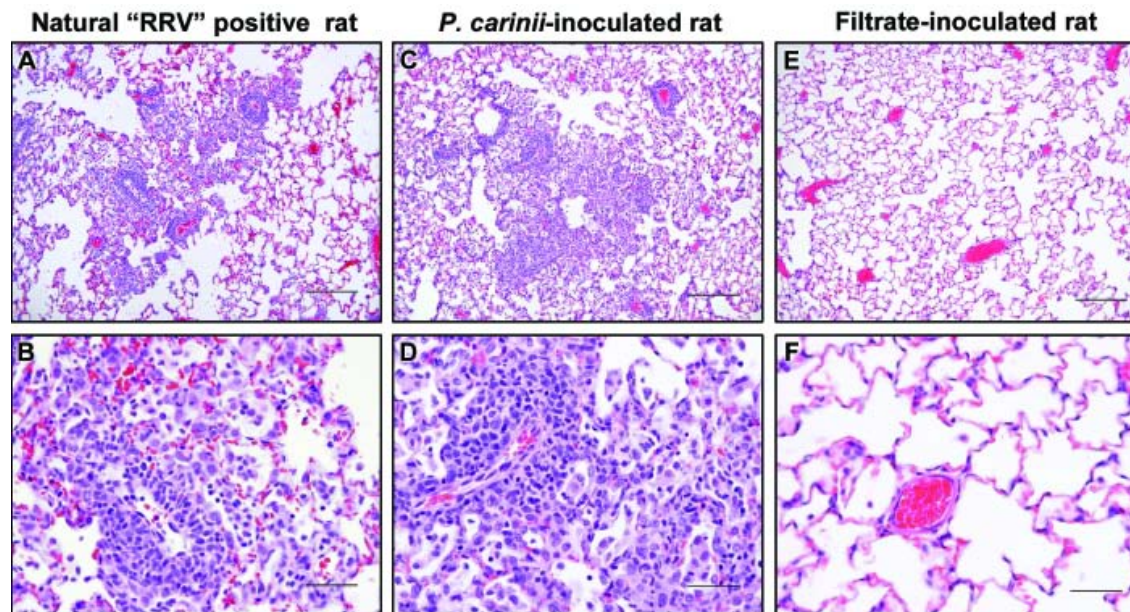
- A health monitoring program should be a dynamic entity of any laboratory animal program. Health status can directly impact research results and should be protected through routine monitoring of animals within the colony.
- The design should include appropriate sample size and organisms monitored, as well as reliable testing methodology.
- Procedures should be in place to reduce introduction of pathogens, including appropriate biosecurity measures and approval of animal sources.
- Formulas exist to calculate proper sample size based on known organism prevalence and colony size, but what organisms should be monitored? How does one account for emerging pathogens?

Novel organisms – Murine Norovirus⁵

- Murine Norovirus
 - Discovered in 2003 in immunocompromised mouse colony; clinical symptoms elicited in immunodeficient mice
 - Immunocompetent mice can be infected but may not display clinical symptoms

Novel organisms – *Pneumocystis carinii*⁶

- *Pneumocystis carinii*
 - Determined in 2010 as causative agent of idiopathic lung lesions in rats
 - Example of known disease with no causative agent identified; now easily monitored within colonies



Sample testing size

- ILAR formula (1976)
 - Statistical analysis
 - Infectivity of organism
 - Production and husbandry procedures
- Number of animals sampled depends on nature of infection and husbandry and caging
- As prevalence of infection decreases, sample size required to detect the infection increases



Sample testing size

Table 1 Calculation of the number of animals to be monitored

Diseases with an infection rate of 50% or more (Sendai, MHV) require far fewer animals to detect their presence than diseases with low infection rates.

Assumptions

1. Both sexes are infected at the same rate
2. Population size > 100 animals
3. Random sampling
4. Random distribution of infection

The sample size is calculated from the following formula:

$$\frac{\log 0.05}{\log N} = \text{Sample size}$$

N = percentage of non-infected animals

0.05 = 95% confidence level

Relation of sample size to prevalence rate

Suspected prevalence rate (%)	Sample sizes at different confidence levels		
	95%	99%	99.9%
10	29	44	66
20	14	21	31
30	10	13	20
40	6	10	14
50	5	7	10

Example: 10 animals should be monitored to detect at least one positive animal if the suspected prevalence rate of an infection is 30% (confidence level: 95%)

Nicklas et al. 2002

Sentinel Type

- Contact Sentinels
 - Most reliable
 - Susceptibility to organisms can be strain specific

- Dirty Bedding Sentinels
 - Susceptibility to organisms can be strain specific
 - Many organisms do not transmit well via soiled bedding

- Colony resident
 - Randomly selected
 - Represent true susceptibility of population (if immunocompetent)



Other sample types

- Tissue samples
- Blood samples
- Fecal samples
- Environmental samples



What organisms to include

- FELASA recommendations
- Vendor health reports
- Organisms within the research scope
 - May desire animals negative for organism being studied



Testing Methodologies - Serology

Pros

- Inexpensive
- Easy to collect blood sample
- Can assay multiple organisms, multiple antigens
- Highly sensitive and specific

Cons

- Can take several weeks for seroconversion
- Does not indicate if infection is active or historical
- False positives can occur with older animals



Testing Methodologies - PCR

Pros

- Highly sensitive and specific
- Multiplex approach possible
- Detect active infection or environmental contamination
- Use multiple sample types

Cons

- High cost
- Short duration of shedding may necessitate frequent sampling
- DNA may be degraded
- Susceptible to contamination



Testing Methodologies - Microbiology

Pros

- Relatively low cost
- Tests for wide range of organisms
- Detects active infection

Cons

- Susceptible to contamination
- Requires careful sample handling
- Fastidious organisms can be difficult to detect
- Variable characteristics of some organisms



Testing Methodologies - Parasitology

Pros

- Relatively low cost
- Tests for wide range of organisms
- Detects active infection



Cons

- Relatively insensitive
- Variable shedding of some organisms
- Dependent on skill of technician



Biosecurity

- Correlates to success of health monitoring program
 - Entry procedures
 - Sterilization of supplies
 - Cage changing
 - Animal procedures
- Proper review of animal sources entering facility
 - Quarantine if needed
- Implement biosecurity auditing program within one's facility to ensure compliance and health of animals

Conclusion

- Proper sampling size, sentinel selection, and test methodology lead to protection of research results by reducing variation due to health status
- Novel organisms may be more quickly discovered and therefore controlled or eradicated through proper health monitoring program implementation
- Health monitoring is an integral component of proper animal research and measures should be taken to ensure accuracy and integrity of the health monitoring program.

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